

**Veterinary parasitology and entomology pdf**

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Authors who submit papers based on local data will need to indicate why their paper is relevant to a broader readership. Or they can submit to the journal's companion title, Veterinary Parasitology: Regional Studies and Reports, which welcomes manuscripts with a regional focus. Parasitological studies on laboratory animals fall within the scope of Veterinary Parasitology only if they provide a reasonably close model of a disease of domestic animals. Additionally the journal will consider papers relating to wildlife species where they may act as disease reservoirs to domestic animals, or as a zoonotic reservoir. Case studies considered to be unique or of specific interest to the journal, will also be considered on occasions at the Editors' discretion. Papers dealing exclusively with the taxonomy of parasites do not fall within the scope of the journal. Studies on rickettsial disease organisms (Ehrlichia, Anaplasma, Eperythrozoon) will be considered for publication in Veterinary Parasitology, but only if the paper deals with vector transmission of these organisms to domesticated animals, or if zoonotic. Studies on Rickettsia per se will not be accepted. Studies dealing with parasite control by means of natural products, both in vivo and in vitro, fall within the scope of the journal, but only if well documented and with therapeutically relevant minimum inhibitory concentrations of the active compound(s) being clearly demonstrated. Circumstances relating to animal experimentation must meet the International Guiding Principles for Biomedical Research Involving Animals as issued by the Council for International Organizations of Medical Sciences. (Obtainable from: Executive Secretary C.I.O.M.S., c/o W.H.O., Via Appia, CH-1211 Geneva 27, Switzerland.) Manuscripts reporting meta-analyses and systematic reviews that follow PRISMA or MOOSE reporting guidelines will receive consideration only if they go beyond reporting parasite prevalence and provide a description and analysis of factors and mechanisms associated with the reported data. Submit your paper! The Impact Factor of this journal is 2.821, ranking it 23 out of 144 in Veterinary SciencesWith this journal indexed in 11 international databases, your published article can be read and cited by researchers worldwideView articlesView historical data and other metrics on Journal Insights. 50 editors and editorial board members in 20 countries/regionsUSUnited States of America (8)and 17 moreEditor-in-Chief Andy Greer, BAgRSc(Hons), PhDEditor-in-Chief Martin K. Nielsen, DVM, Dipl. EVPCEditor-in-Chief Michael P. Reichel, VS, DVPH, DTV, MVSc, MBA, Dr med vet, PhD, FACVSc, FRCVS, Dipl ECVPHEditor-in-Chief Theo de Waal, BVSc, PhDView editorial boardJournalFinder uses smart search technology and field-of-research specific vocabularies to match your manuscript to Elsevier journals. Simply insert your title and abstract and select the appropriate field-of-research for the best results. Fill in details For the journal, see Veterinary Parasitology (journal). Veterinary parasitology is the study of animal parasites, especially relationships between parasites and animal hosts. Parasites of domestic animals, (livestock and pet animals), as well as wildlife animals are considered. Veterinary parasitologists study the genesis and development of parasites in animal hosts, as well as the taxonomy and systematics of parasites, including the morphology, life cycles, and living needs of parasites in the environment and in animal hosts. Using a variety of research methods, they diagnose, treat, and prevent animal parasites. Data obtained from parasitological research in animals helps in veterinary practice and improves animal breeding. The major goal of veterinary parasitology is to protect animals and improve their health, but because a number of animal parasites are transmitted to humans, veterinary parasitology is also important for public health.[1][2][3] Diagnostic methods Various methods are used to identify parasites in animals, using feces, blood, and tissue samples from the host animal.[2] Coprological examinations involve examining the feces of animals to identify and count parasite eggs. Some common methods include fecal flotation and sedimentation to separate eggs from fecal matter. Others include the McMaster method, which uses a special two-chamber slide that allows parasite eggs to be more clearly visible and easily counted. It is most commonly used to monitor parasites in horses and other grazing and livestock animals. The Baermann method is similar but requires more specialized equipment and more time and is typically used to diagnose lungworm and threadworm.[4] Haematological examinations involve examining the blood of animals to determine the presence of parasites. Blood parasites tend to inhabit the erythrocytes or white blood cells and are most likely to be detected during the acute phase of infection. Veterinary parasitologists use blood smears, which involve placing a drop of blood onto a slide and spreading it over the surface in a thin film in order to examine it under a microscope. The blood is stained with a dye in order for the cells to be easily distinguished.[5] Histopathological examinations involve examining tissue samples from animals. A small slice of the organ suspected of being infected by parasites is mounted on a slide, stained, and examined under a microscope. Though not technically considered a histopathological technique, skin scraping – which involves taking a small sample of the epidermal cells of a dog, cat, or other household pet – is commonly used to detect the presence of mites.[4] Immunological examinations, such as indirect immunofluorescence, ELISA, Immunoblotting (Western blot), and Complement fixation test are methods of identifying different kinds of parasites by detecting the presence of their antigens on or within the parasite itself. These diagnostic methods are used in conjunction with coprological examinations for more specific identification of different parasite species in fecal samples.[5] Molecular biological methods involve studying the DNA of the parasite in order to identify it. PCR and RFLP are used to detect and amplify parasite DNA found in the feces, blood, or tissue of the host. These techniques are very sensitive, which is useful for diagnosing parasites even when they are present in very low numbers; they are also useful for identifying parasites not only in large animal hosts but smaller insect vectors.[6] Divisions of veterinary parasitology Veterinary protozoology is focused on protozoa with veterinary relevance. Examples of protozoan parasites: Babesia divergens Balamutidium coll Besnoitia besnoiti Cryptosporidium parvum Eimeria acervulina Eimeria tenella Giardia lamblia (also known as Giardia duodenalis) Hammondia hammondi Histomonas meleagridis Isospora canis Leishmania donovani Leishmania infantum Neospora caninum Toxoplasma gondii Trichomonas gallinae Trichomonas foetus Trypanosoma equiperdum Veterinary helminthology Veterinary helminthology is focused on veterinary important helminth parasites, for example: Ancylostoma caninum Ancylostoma ceylanicum Ancylostoma duodenale Ascaris suum Dicrocoelium dendriticum Dictyocaulus viviparus Dipylidium caninum Echinococcus granulosus Fasciola hepatica Fascioloides magna Habronema species Haemonchus contortus Metastrombus Muellerius capillaris Ostertagia ostertagi Paragonimus westermani Schistosoma bovis Strongylidae species Strongylus vulgaris Syngamus trachea (Gapeworm) Taenia pisiformis Taenia saginata Taenia solium Toxascaris leonina Toxocara canis Toxocara cati Trichinella spiralis Trichobilharzia regenti Trichostrongylus species Trichuris suis Trichurus vulpis Veterinary entomology (arachnometiology) Veterinary entomology is focused on important arachnids, insects, and crustaceans. Some examples include: Caligus species Cimex pilosellae Cimex lectularius Culex pipiens Culicoides imicola Demodex bovis Dermacentor reticulatus Gasiphilus intestinalis Haematocheilus irritans Hypoderma bovis Ixodes ricinus Knemidocoptes mite (causing the disease scaly leg) Leucophtheirus salmonis (sea louse) Lucifer sericata Musca domestica Nosema apis Nosema cati Oocystis ovis Otodectes cynotis Phlebotomus species Psoroptes ovis Pulicaria irritans Rhinicephalus sanguineus Sampsoniella coquilletti Tabanus atratus Triatoma species Ctenocephalides felis References ^ Jurásek V., Dubinský P., 1993. Veterinárná parazitológia. Príroda a.s., Bratislava, 382 pp. ^ a b Kauffman, J., 1996. Parasitic infections of domestic animals: a diagnostic manual. Birkhäuser, Berlin, 423 pp. ^ Elsheikha, H.M., Khan, N.A. (editor), 2011. Essentials of Veterinary Parasitology. Cisterc Academic Press, ISBN 978-1-904455-79-4. {cite book}}; author = "Jurásek V., Dubinský P.", title = "Veterinárná parazitológia", publisher = "Príroda a.s.", year = "1993", pages = "382", url = "http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3130722/", language = "Slovak", note = "helpCS1 maint: multiple names: authors list (link)" ^ a b Leland S., Shiroki, R., & Mendel (22 May 2009). Pathology & Parasitology for Veterinary Technicians. Cengage Learning. pp. 237–239. ISBN 978-1-4354-3855-2. Retrieved 11 September 2011. ^ a b A. Zajac; Gary A. Conboy; American Association of Veterinary Parasitologists (24 March 2006). Veterinary clinical parasitology. Wiley-Blackwell. p. 162. ISBN 978-0-8138-1734-7. Retrieved 11 September 2011. ^ John E. Hyde (1993). Protocols in molecular parasitology. Humana Press. pp. 213–214. ISBN 978-0-89603-239-2. Retrieved 11 September 2011. External links World Association for Advancement of Veterinary Parasitology Guide to Veterinary Diagnostic Parasitology American Association of Veterinary Parasitologists Retrieved from "https://en.wikipedia.org/w/index.php?title=Veterinary\_Parasitology&oldid=900000000"}

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